Revisional study on African *Apophylia*. Part 5
(Coleoptera: Chrysomelidae: Galerucinae)

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**ABSTRACT.** The fifth contribution to the knowledge of African *Apophylia* THOMSON, 1858 based on the study of type materials is presented. *Apophylia grobbelaarae* n. sp. (from RSA and Zimbabwe), *A. haladai* n. sp. (from Guinea), *A. lindae* n. sp. (from Namibia), *A. marketae* n. sp. (from Tanzania, Zambia and Zimbabwe), *A. dellacasai* n. sp. (from Egypt and Oman) and *A. lesnei aethiopica* n. ssp. (from Ethiopia) are described. *Apophylia similis* WEISE, 1909 and *A. disconotata* PIC, 1947 are redescribed. The lectotypes are designated for *Apophylia lata* PIC, 1945, *A. marginipennis* WEISE, 1912, *A. jeanneli* LABOISSIÈRE, 1921, *A. lesnei* LABOISSIÈRE, 1922, *Malaxia aurolimbata* ALLARD, 1888 and *M. marshalli* JACOBY, 1897. Following new synonyms are proposed: *A. lata* PIC, 1945 = *A. clavareaui* LABOISSIÈRE, 1940; *A. aurolimbata* (ALLARD, 1888) = *A. femorata* (JACOBY, 1895). *Apophylia scutellaris* ALLARD, 1889 is transferred to the genus *Chapuisia* (comb. nov.) and *Apophylia aeneipennis* (ILLIGER, 1800) to the genus *Taumacera* (comb. nov.). A new name *Chapuisia weisei* nom. nov. is proposed for *Chapuisia scutellaris* WEISE, 1927 nec *Chapuisia scutellaris* (ALLARD, 1889). Male genitalia of most of the *Apophylia* species studied are figured.

Key words: entomology, taxonomy, lectotype designation, synonymy, Coleoptera, Chrysomelidae, Galerucinae, *Apophylia*, Afrotropical Region

The following abbreviations identify the collections housing the examined material:

BMNH - United Kingdom, London, The Natural History Museum (Sharon SHUTE);
CIUC - Italy, Calci, Centro Interdipartimentale dell’Università, Museo di Storia Naturale e del Territorio (Marco DELLACASA);
ISNB - Belgium, Brussels, Institut Royal des Sciences Naturelles de Belgique (Didier DRUHMAND, Marcel CLUDTS);
JBCB - Czech Republic, Brno, Jan BEZDĚK collection;
JVCJ - Czech Republic, Jirkov, Jiří VOŘÍŠEK collection;
MCSN - Italy, Genova, Museo Civico di Storia Naturale „Giacomo Doria” (Roberto POGGI);
MCST - Italy, Trieste, Museo Civico di Storia Naturale (Andrea COLLA)
MCZC - USA, Massachusetts, Cambridge, Museum of Comparative Zoology (Phillip D. PERKINS);
MNHN - France, Paris, Muséum National d’Histoire Naturelle (Nicole BERTI);
MRAC - Belgium, Tervuren, Musée Royal de l’Afrique Centrale (Mark DE MEYER);
NHMB - Switzerland, Basel, Naturhistorisches Museum (Eva SPRECHER-UEBERSAX, Michel BRANCUCCI);
NMPC – National Museum, Praha, Czech Republic (Jiří HAJEK);
RBCN - Netherlands, Nieuwegein, Ron BEENEN collection;
SANC - South Africa, Pretoria, South African National Collection of Insects (Elizabeth GROBBELAAR);
SMNS - Germany, Stuttgart, Staatliches Museum für Naturkunde (Wolfgang SCHWALLER);
TMSA - South Africa, Gauteng, Pretoria, Transvaal Museum (Ruth MULLER);
USNM - USA, Washington D.C., National Museum of Natural History (Alexander KONSTANTINOV);
ZMHB - Germany, Berlin, Museum für Naturkunde der Humboldt-Universität (Johannes FRISCH, Joachim WILLERS).

Exact label data are cited for all type specimens; a double slash (//) divides data on different labels. Type localities are cited in the original spelling. Other comments and remarks are placed in square brackets: [p] – preceding data are printed; [h] – the same, but handwritten; [w] - white label; x/y - number of males/number of females. The lectotypes and paralectotypes are designated in order to preserve stability of nomenclature in this group, according to the Article 74.7.3 of the Code (ICZN 1999).

**Taumacera aeneipennis (ILLIGER, 1800), comb. nov.**

*Galleruca aeneipennis* ILLIGER, 1800: Arch. Zool. Zoot. 1:134 (Type locality: Africa)  
*Apophylia aeneipennis* GEMMINGER & HAROLD 1876: 3569; ALLARD 1889: 71; WEISE 1924: 183; WILCOX 1971: 142.

**Type material examined**

COMMENTS

Galleruca aeneipennis was described based on one female deposited in ZMHB. In their catalogue GEMMINGER & HAROLD (1876) listed it under the genus Apophylia with some doubts. Also ALLARD (1889) considered its position within Apophylia doubtful, but he did not examine the holotype and did not suggest any generic replacement.

Galleruca aeneipennis seems to be closely related to African species of Platyxantha BALY, 1864. Recently, REID (1999) revised the generic positions of Asiatic genera closely related to Taumacera THUNBERG, 1814 and synonymized Platyxantha with Taumacera. The African Platyxantha species were placed in Taumacera with some doubts. I decided to follow the REID’s conception and tentatively suggest the transfer of Galleruca aeneipennis to Taumacera. However, a detailed study of African members of previous Platyxantha and allied genera is highly welcome.

Apophylia aurolimbata (ALLARD, 1888)

Malaxia aurolimbata ALLARD, 1888: Ann. Soc. Ent. Fr. (6)8: 332 (Type locality: Natal); ALLARD, 1889: 80 (sep. 15).
Apophylia aurolimbata: LABOISSIÈRE 1922b: 243 (key) (sep. 155); WEISE 1924: 183; WILCOX 1971: 142.
Apophylia femorata: LABOISSIÈRE 1922b: 245 (key) (sep. 157); WEISE 1924: 183; LABOISSIÈRE 1940: 14 (key); WILCOX 1971: 144; BEZDĚK 2004: 99.

TYPE MATERIAL EXAMINED

Malaxia aurolimbata
Lectotype (female), designated here, and 1 paralectotype (female), labelled: “Natal [w, h] // Ex-Musæo E. ALLARD 1899 [w, p]“ (in MNHN). The specimens are provided with one red label: „LECTOTYPUS [or PARALECTOTYPUS], Malaxia aurolimbata Allard, 1888, des. J. Bezděk 2004”.

Malaxia femorata

ADDITIONAL MATERIAL EXAMINED

RSA: Zululand, Hluhluwe Game Res., 28°05´S 32°04´E, 27.xi.1992, Endrödy-Younga leg. (0/2 in TMSA); same data, 18.xi.1992 (1/0 in TMSA); Transkei, Dwesa, 32°17´S 28°51´E, 28.ii.1985, Endrödy-Younga leg. (0/1 in TMSA); Algoa Bay, Brauns leg. (0/1 in TMSA); Bashee Bridge, 3.xii.1956, R. M. Martin leg.
(13/8 in TMSA); Port St. Johns, 24.-30.xi.1956 (0/1 in TMSA); Port St. John, x.1923, R. E. Turner leg. (1/0 in TMSA); 33 km S Fort Beaufort, 33°02’S 26°39’E, collected on *Ehretia rigida* (Boraginaceae), 23.xi.1988, B. Grobbelaar leg. (3/10 in SANC); Cintsa Mouth, 32°49’S 28°07’E, 26.xi.1988, R. Oberprieler leg. (1/0 in SANC); Andries Vosloo Kudu reserve near Grahamstown, 33°07’S 26°38’E, 30.xi.1983, R. Oberprieler leg. (1/3 in SANC); Transvaal, Barberton, 25°48’S 31°03’E, iii.1979, C. G. E. Moolman leg. (1/1 in SANC); Nico Malan Pass, 5 km NE Seymour, 1400 m, 32°30’S 26°50’E, 25.xi.1988, B. Grobbelaar leg. (1/1 in SANC); Natal, Richmond, Mahlaleen river, xii.1959, E. Haaf leg. (4/8 in NHMB – Frey coll.); Natal, Richmond distr., Umkoma riv. valley, 1.xii.1956 (1/0 in MRAC); Natal, Malvern, 24.ii.1902 (1/0 in USNM); Caffrarie, Schaum leg. (2/0 in DEI).

Aedeagus as in Fig. 1.

**BIONOMY**

Several specimens were collected on *Ehretia rigida* (Boraginaceae).

**DISTRIBUTION**

RSA.

**COMMENTS**

*Apophylia aurolimbata* was described based on two females from Natal deposited now in MNHN. The type series of *A. femorata* consists of males only. The specimens of *A. femorata* are nothing but males of *A. aurolimbata*. Three specimens deposited in TMSA and NHMB and published as females of *A. femorata* by BEZDÍK (2004) are strongly damaged and proved to be males. The sexual dimorphism is very remarkable - males have black pronotum and femora with two basal thirds black, females have yellow pronotum with three black spots and yellow legs with black spots near femoral base. Similar sexual dimorphism is known also in other *Apophylia* species, such as *A. maynei* LABOISSIÈRE, 1922, *A. pulchella* BRYANT, 1952, *A. zoiai* BEZDÍK, 2005, *A. hanka* BEZDÍK, 2005 and, partially, *A. grobbelaarae* n. sp.

The males of *A. aurolimbata* are similar to the dark males of *A. grobbelaarae* n. sp., but differ in the black underside of head (yellow in *A. grobbelaarae* n. sp.) and in the structure of aedeagus (Figs 1 and 13). The females of *A. aurolimbata* are similar to females of *A. clavareaui* and *A. grobbelaarae* n. sp., but these species have yellow underside of head (black in *A. aurolimbata*). Moreover, *A. clavareaui* has also yellow frontal tubercles (black in *A. aurolimbata*).

*Apophylia clavareaui* LABOISSIÈRE, 1940


*Apophylia clavareaui*: WILCOX 1971: 143.
**Type material examined**

*Apophylia clavareaui*


*Apophylia lata*


**Additional material examined**

BOTSWANA: Kanye, xii.1955, Zumpt leg. (1/1 in NHMB – Frey coll.); Tsane, xii.1954, Zumpt leg. (0/1 in NHMB – Frey coll.); 80 km N Palapye, 29.xii.1972, E. Holm & D. Paterson leg. (1/0 in SANC); NAMIBIA: Regenstein, 15 miles SSW of Windhoek, 8.ii.1972 (0/1 in BMNH); Windhoek, 22°34′S 17°05′E, 12.iii.1974, R. Oberprieler leg. (0/1 in SANC); RSA: Natal, Zululand, Mtubatuba, 24.-25.iii.1968, P. J. Spangler leg. (30/5 in USNM); Natal, Sodwana Bay Park, 27°32′S 32°41′E, 9.-11.xi.1986, D. D. Hotman & A. Nel leg. (1/0 in SANC); Natal, St. Lucia Estuary, 28°17′S 32°25′E, 26.ii.1989, E. Grobbelaar & E. v. d. Linde leg. (0/1 in SANC); Natal, Kosi Bay, Banga Nek, 27°00′S 32°53′E, 50 m, 11.ii.1990, E. Grobbelaar leg. (1/1 in SANC); Natal, Tsikingo, ii. 1896 (0/1 in BMNH); Natal, Malvern, 1912, G. A. K. Marshall leg. (2/0 in BMNH); Kwazulu, Lake Sibaya, E shore, 27°22′S 32°43′E, 18.-20.i.1981, R. Oberprieler leg. (4/3 in SANC); Kwazulu-Natal, False Bay, Lake St. Lucia, E shore, 27°22′S 32°43′E, 2.xi.1991, P. E. Reavell leg. (0/1 in SANC); Kwazulu-Natal, Sodwana Bay N. P., 27°37′S 32°41′E, 20.xi.1995, F. Koch leg. (2/5 in ZMHB); same data, 30.i.-1.ii.1994, U. Göllner leg. (0/4 in ZMHB); Kwazulu-Natal, N’dumu Game
Reserve, 26°55´S 32°19´E, 25.-27.xi.1995, F. Koch leg. (1/5 in ZMHB); Kwazulu-Natal, near Lake Nhlabane, 25 km NE Richards Bay, 28°38´S 32°16´E, 30.iii.1991, M. Vogt leg. (1/0 in SANC); same data, 14.xi.1991 (0/2 in SANC); Northern prov., Kommandonek, 25°45´S 27°47´E, 1.xii.1995, C. L. Bellamy leg. (2/7 in TMSA); Northern prov., Silkaatsneck, 25°40´S 27°55´E, 30.xi.1995, C. L. Bellamy leg. (0/2 in TMSA); Pretoria Fountains, 25°57´S 28°12´E, 19.xii.1985, Endrödy-Younga leg. (0/1 in TMSA); Pretoria, xii.1979, A. P. du Toit leg. (1/0 in SANC); Pretoria, xi.1978, M. Edwards leg. (0/1 in SANC); Pretoria, swept garden PPRI, 26.xi.1994, K. W. R. Zwart leg. (4/1 in SANC); Transvaal, 15 km E of Pretoria, 10.xi.1983, C. L. Bellamy leg. (2/0 in TMSA); Transvaal, Baviaanskloof, SW of Potgietersrus, 24°18´S 28°55´E, 1.i.1993, E. Grobbelaar leg. (0/1 in SANC); Pretoria, Springbokparkie, 25°47´S 28°17´E, 13.iii.1997, M. Botha leg. (0/1 in SANC); Pretoria, Meintjies Kop, 15.xii.1964, M. Hoffmann leg. (0/2 in SANC); Transvaal, Percy Fyfe Nat. Res., 24°03´S 29°09´E, 10.-12.iii.1980, C. Kok leg. (1/0 in SANC); Transvaal, Ellisras distr., D’Nyala Nat. Res., 23°45´S 27°49´E, 8.-12.xii.1989, C. D. Eardley leg. (1/0 in SANC); Transvaal, Lapalala Nat. Res., 23°51´S 28°17´E, 19.ii.1994, R. Oberprieler leg. (0/1 in SANC); Transvaal, Sterkriver near Naboomspruit, 24°17´S 28°48´E, 1.iii.1991, V. M. Uys leg. (0/1 in SANC); Transvaal, Rooiwal, SW of Potgietersrus, 24°14´S 28°49´E, 1.ii.1993, E. Grobbelaar leg. (0/1 in SANC); Zululand, Hluhluwe Game Reserve, 28°05´S 32°04´E, 27.xi.1992, Endrödy-Younga leg. (0/1 in TMSA); Cape of Good Hope (1/0 in BMNH); Limpopo, 5 km NNE Thengwe, 700 m, 22°40´S 30°34´E, collected from *Ehretia rigida* (Boraginaceae), 6.ii.1994, E. Grobbelaar leg. (5/9 in SANC); Venda Thengwe, 22°40´S 30°34´E, 650 m, 7.ii.1994, E. Grobbelaar leg. (0/1 in SANC); Limpopo, Mogol Nature Reserve, Ellisras District, 23°58´S 27°54´E, 21.xii.1987, E. Grobbelaar leg. (0/2 in SANC); Naboomspruit, 24°31´S 28°43´E, 27.-29.ii.2004, S. J. Burger & C. A. Strange leg. (0/3 in SANC); Transvaal, Farm Kuleni, Hluhluwe distr., 27°54´S 32°22´E, 50 m, 12.-14.ii.1990, C. D. Eardley leg. (0/1 in SANC); same data, 13.-14.ii.1990, N. Verheijen leg. (1/0 in SANC); Natal, Durban, 1904, J. P. Cregoe leg. (0/3 in BMNH); Horns Nek, 10.iii.1966, A. L. Capener leg. (3/0 in SANC); Pietersburg, 7.xii.1965, A. L. Capener leg. (0/1 in SANC); North Cape, 75 km ENE of Kuruman near Lykso, 27°25´S 24°07´E, 1.-6.iv.2002, E. Holm & H. Gebhardt leg. (1/0 in SANC); Mtunzini, xii.1979, R. Oberprieler leg. (0/1 in SANC); Bultfontein, ii.1921, H. K. Munro leg. (0/1 in SANC); Chuniespoort, 6.xii.1965, P. Palaitseas leg. (0/1 in SANC); ZIMBABWE: Bulawayo, 19.xii.1924, R. H. R. Stevenson leg. (0/1 in TMSA); Bulawayo, 1903, F. Brooks leg. (1/1 in BMNH).

Aedeagus as in Figs 2-3.

Bionomy

Several specimens were collected on *Ehretia rigida* (Boraginaceae).

Distribution

Botswana, Mozambique, Namibia, RSA, and Zimbabwe.
COMMENTS

*A. clavareauui* is similar to *A. lindae* n. sp., to females of *A. aurolimbata* and to females and paler males of *A. grobbleaarae* n. sp. The females of *A. aurolimbata* differ in the black underside of head and frontal tubercles (yellow in *A. clavareauui*). The females and the paler males of *A. grobbleaarae* n. sp. have black frontal tubercles (yellow in *A. clavareauui*). *A. lindae* n. sp. differs in the completely yellow postgenae (bicolorous in *A. clavareauui*) and somehow robuster antennomeres than *A. clavareauui*. Moreover, all mentioned species can be easily distinguished by the structure of aedeagi (Figs 1-3, 13, 15).

The type series of *A. clavareauui* (3 females) was compared with two syntypes of *A. lata* (male and female, male is designated here as lectotype) and both species are evidently conspecific, thus *A. lata* is considered a new synonym of *A. clavareauui*. Moreover, the paratype of *A. clavareauui* from Mozambique bears an identical label as both types of *A. lata*. The aedeagus of the lectotype of *A. lata* is weakly sclerotized and damaged in apical part (compare Figs 2 and 3).

Apophylia disconotata Pic, 1947

Apophylia disconotata Pic, 1947: Divers. Ent. II: 4 (Type locality: Abyssinie); Wilcox 1971: 143.

Type material examined

Additional material examined
ETHIOPIA: Abyssinia, Raffray leg. (0/1 in MNHN); Ilubabor prov., 15 km NW of Chora, 1600 m, vi. 1973, G. de Rougemont leg. (1/2 in MRAC).

Redescription
Body length of male 9.00 mm; of females 9.25-9.75 mm.


Labrum transverse, laterally covered with several pale setae, anterior margin sinuate. Anterior part of head lustrous, sparsely covered with pale setae. Interantennal space with small feeble depression. Frontal tubercles small, subtriangular, covered with microsculpture, semiopaque. Vertex dull, densely covered with small confused punctures and short pale hairs. Antennae filiform, length ratio of antennomeres 1 to 5: 25-11-24-34-30 (rest missing).

Pronotum transverse, 1.70 times broader than it is long, widest at the first third, slightly narrowed anteriad and posteriad, dull, densely covered with small punctures and pale hairs. Surface with two feeble depressions laterally connected with indistinct very feebly impressed ridge. Posterior angles surrounded by very small feeble depressions. Anterior margin slightly concave, posterior margin nearly straight, lateral margins slightly rounded. Anterior and posterior margins thinly bordered, lateral margins indistinctly bordered. Anterior angles nearly rectangular, posterior angles obtusely angulate, all angles with distinct teeth bearing long pale setae.

Scutellum short, subtriangular, with small dense punctures and short pale hairs, semiopaque.

Elytra parallel, dull. Humeral calli well developed. Elytral surface very densely covered with small confused punctures and short pale hairs. Epipleura distinct, gradually narrowed to apex.

Macropterous.

Ventral surface lustrous, finely punctured and covered with pale hairs. Last visible sternite with very deep semicircular incision.
First tarsomeres enlarged, basimetatarsomere 1.35 times as long as two following metatarsomeres combined (Fig. 9).

Female: Pronotum bicolorous, yellow with large central black spot and very small spots in all angles around setigerous pores. Abdomen black, last two ventrites brown to yellow, pygidium yellow. First tarsomeres not enlarged. Last visible sternite with small sharp incision. Claws appendiculate.
Aedeagus as in Fig. 6.

**Distribution**
Ethiopia.

**Diagnosis**
One of the largest *Apophylia* species. The male of *A. disconotata* resembles the males of *A. maynei* Laboissière, 1922, *A. pulchella* Bryant, 1952, *A. zoiai* Bezděk, 2005 and *A. hanka* Bezděk, 2005 but differs in the structure of tarsi. All basitarsomeres are strongly enlarged in *A. disconotata*, while the other mentioned species have basimetatarsomere slightly enlarged.

Due to the coloration of pronotum, the females are very similar to the female of *A. trapezicollis* Laboissière, 1940 (only female holotype known). Females of both species differ in the length ratio of antennomeres 2 and 3. Antennomere 3 is 2.2 times longer in the female of *A. disconotata*, but only 1.8 times in the female of *A. trapezicollis*. The central spot on pronotum is larger in the female of *A. disconotata*.

**Comments**
*A. disconotata* was described based on one female. In unidentified material from MRAC I have found short series including one male. An additional female was found also in MNHN. Because the male was not known, the redescription of this species is presented.

*Apophylia jeanneli* Laboissière, 1921

*Apophylia jeanneli* Laboissière, 1921: Bull. Soc. Ent. Fr., 1921: 8 (Type locality: Bura); Laboissière 1922b: 240 (key) (sep. 152); Laboissière 1925: 57; Laboissière 1929: 340.


**Type Material Examined**
JEANNEL Mars 1912 – 1050\textsuperscript{m} – St. 61 [w, p] // Apophylia Jeanneli m [h] V. Laboissière – Dét. [p] 1921 [w, h] // Para-type [orange label, p]“ (in ISNB). The specimens are provided with one red label: „LECTOTYPUS [or PARALECTOTYPUS], Apophylia Jeanneli Laboissière, 1921, des. J. Bezděk 2004”.

ADDITIONAL MATERIAL EXAMINED
KENYA: Eastern Nyambeni Hills, Ngaja Forest, 0\degree19’113’’N 38\degree02’609’’E, 1070m, at light, 2.-4.xii.2002, C. Häuser, D. Bartsch & A. Zahm leg. (1/0 in SMNS) Fort Hall, 5.xii.1919, Patrizi leg. (5/18 in MCSN); Fort Hall, xii.1919, Patrizi leg. (3/7 in MCSN); Kisumu, 22.-24.iv.1911, S. A. Neave leg. (1/1 in BMNH); TANZANIA: W, S, SE edge Makata plain, 9.iii\textsuperscript{2}2002, M. Sní ek leg. (1/0 in JBCB); Handeni, Makinda env., 14.iii.2002, M. Sní ek leg. (1/0 in JBCB); Handeni, 350 m, 25.-27.iv.1957, P. Basilewsky & N. Leleup leg. (1/0 in MRAC); UGANDA: Semliki plains, near S shore of Lake Albert, 25.-27.xi.1911, S. A. Neave leg. (1/1 in BMNH).

Aedeagus as in Fig. 10.

DISTRIBUTION
Kenya, Tanzania.

COMMENTS
A species with yellow vertex with central black spot. Externally, \textit{A. jeanneli} is very similar to \textit{A. marshalli} (JACOBY, 1897) and \textit{A. marketae} n. sp. Exact identification is possible only based on the structure of aedeagi of all three species (Figs 10-11, 16).

LABOISSIÈRE (1921) did not specify the number of available specimens, but mentioned sexual dimorphism. I have found only two type specimens: a male deposited in MNHN (designated here as lectotype) and a female in ISNB. The aedeagus of the lectotype is very weakly sclerotized, the drawing is based on another specimen.

\textit{Apophylia lesnei} LABOISSIÈRE, 1922


\textit{Apophylia lesnei}: WILCOX 1971: 145.

TYPE MATERIAL EXAMINED
type [orange label, p]“ (in ISNB). The specimens are provided with one red label: „LECTOTYPUS [or PARALECTOTYPUS], Apophylia Lesnei Laboissière, 1922, des. J. Bezděk 2004”.

ADDITIONAL MATERIAL EXAMINED
KENYA: Kerio river, vi.1914, Dr. Bayer leg. (1/0 in MRAC).
Aedeagus as in Fig. 4.

DISTRIBUTION
Kenya.

COMMENTS
A species with yellow head with black vertex and yellow pronotum with three black spots (median and two lateral). *A. lesnei* differs from its congeners in the remarkable aedeagus (Fig. 4) and in enlarged basitarsomerones of all legs in male (basimetatarsomere as in Fig. 7). *A. lesnei aethiopica* n. ssp. differs from the nominate subspecies in strongly enlarged basitarsomerones (Fig. 8).

*Apophylia marginicollis* LABOISSIÈRE, 1940


TYPE MATERIAL EXAMINED

**Distribution**

Congo.

**Comments**

The type series consists of 6 females. The exact identification of this species is very problematic due to unknown males. With the coloration of pronotum (yellow with black lateral spots, without median spot) it seems to be closely related to *A. vicina* (unfortunately known also only from female holotype). Both species differ only in the colour of frontal tubercles, which are black in *A. marginicollis*, while yellow in *A. vicina*.

*Apophylia marginipennis* Weise, 1912

*Apophylia marginipennis* Weise, 1912: Deutsche Zentr.-Afr.-Exp. 4(7): 149 (NW von Beni; Mawambi am Ituri); Laboissière 1922b: 244 (key), 249 (sep. 156, 161); Weise 1924: 183; Wilcox 1971: 145.

**Type material examined**


**Distribution**

Congo.

**Comments**

The lectotype is characterized by following features: head black with anterior part yellow; frontal tubercles and labrum black; legs yellow, outside profemora with small darkened spot in the middle; pronotum yellow with two small lateral
spots and indistinct median spot; lateral margins of elytra with broad violet band; meso- and metasternum black; abdomen yellow.

Weise (1912) did not specify the number of available specimens. I have found only one type specimen (unfortunately female) which is designated here as lectotype. Exact taxonomic position within the genus *Apophylia* will be cleared when the males are found.

*Apophylia marshalli* (Jacoby, 1897)


**Type material examined**


**Additional material examined**

BOTSWANA: Serowe, 22°25´S 26°44´E, x. 1984, P. Forchhammer leg. (0/1 in SANC); KENYA: Taveta, 750 m, iii.1912, Alluaud and Jeannel leg. (1/3 in ISNB); Eastern Nyambeni Hills, Ngaja Forest, 0°19´113´´N 38°02´609´´E, 1070m, at light, 2.-4.xii.2002, C. Häuser, D. Bartsch & A. Zahm leg. (2/0 in SMNS); Rift Valley Matthews Range, 35 km N of Wamba, 1°10´707´´N 37°18´962´´E, 1300-1400 m, 2.-4.xii.2002, C. Häuser, D. Bartsch & A. Zahm leg. (1/0 in SMNS); Nairobi, vi. 1937, A. F. J. Gedye leg. (1/0 in USNM); RSA: Transvaal, Hans Merensky NR, 20.i.1999, P. Schüle leg. (1/2 in SMNS); Ilbisil env., 50 km N of Namanga, 18.xi.1997, M. Snížek leg. (1/0 in RBCN); 25 km S of Pretoria, Saartjesnek, 16.-18.xii.1997, S. Bílý leg. (1/3 in JVCJ); Transvaal, Vienna Game Farm, Hoedspruit, 500 m, 24°17´S 30°58´E, 17.-18.i.1991, E. Grobbelaar leg. (1/1 in SANC); Transvaal, Hans Merensky Nat. Res., 23°42´S 30°44´E, 23.-25.i.1987, E. Grobbelaar leg. (0/1 in SANC); Transvaal, Swadini, Blydepoort Nat. Res., 24°32´S 30°54´E, 26.-29.i.1987, E. Grobbelaar leg. (0/1 in SANC); Transvaal, Klaserie, 24°33´S 31°01´E, xii.1985, C. H. Scholtz leg. (0/1 in SANC); Transvaal, Kruger Nat. Park, Shingwidi, 350 m, at light, 23°07´S 31°26´E, 6.ii.1988,
E. Grobbelaar leg. (4/7 in SANC); Transvaal, Kruger Nat. Park, Punda Maria, at light, 22°41´S 31°02´E, 7.ii.1988, G. D. Butler leg. (2/0 in SANC); Transvaal, Kruger Nat. Park, Crooke’s Corner near Pafuri, 250 m, at light, 22°23´S 31°15´E, 2.ii.1994, E. Grobbelaar leg. (1/7 in SANC); Natal, near Jozini, 22°38´S 30°24´E, collected on Cordia grandicalyx, 5.-9.ii.1994, E. Grobbelaar leg. (0/2 in SANC); same data, at light (1/1 in SANC); Megaludzo, 23.xi.1969, A. Braack leg. (0/1 in SANC); Zululand, Mkuzi, xii.1945 (0/1 in SANC); SOMALIA: Giuba, Ola Uger, viii.1934, Patrizi leg. (2/6 in MCSN); Giuba, Belet Amin, vii.1934, Patrizi leg. (43/42 in MCSN); TANZANIA: W, S, SE edge Makata plain, 9.iii.2002, M. Snižek leg. (2/0 in JBCB); Kibwesi, Huebner leg. (1/0 in ZMHB); ZAMBIA: South Luangwa N.P., Mfuwe Crocodile Farm, 13°06´S 31°47´E, 450 m, 21.-24.iii.1993, U. Göllner leg. (1/0 in ZMHB); ZIMBABWE: Matobo N. P., 50 km S of Bulawayo, 3.-5.xii.1998, S. Bečvář leg. (1/0 in JBCB); Salisbury, i.1895, G. A. K. Marshall leg. (0/1 in BMNH).

Aedeagus as in Fig. 11.

Host plant: Cordia grandicalyx (Boraginaceae)

**Distribution**


**Diagnosis**

Species with central black spot on yellow vertex and with 3 black spots on pronotum. Very similar to *A. marketae* n. sp. and *A. jeanneli*. All three species can be exactly identified only based on the structure of aedeagus (Figs 10-11, 16).

*Apophylia similis* Weise, 1909


**Type material examined**


**Additional material examined**

TANZANIA: Amani, x.-xii.1905, 900 m, C. Schröder leg. (1/0 in ZMHB, 2/1 in MRAC); Amani, 1.-20.ii.1906, G. Vosseler leg. (0/2 in ZMHB); Amani,
Redescription

Body length of males 4.55-5.00 mm; of females 5.35-6.00 mm (LT 5.40 mm).

Male. Body flattened, parallel, pubescent, semiopaque. Head black with anterior part, mouthparts and the middle of ventral part yellow, frontal tubercles brown to dark brown, mandibles black. Pronotum yellow with three black spots (median and two lateral). Median spot is large and divergent anteriad. Elytra metallic green. Prosternum yellow, mesosternum black, metasternum black with yellow posterior margin. Abdomen yellow to dark brown. Legs yellow, last two tarsomeres infuscate. Antennomeres 1 to 3 yellow, antennomeres 3 and 4 gradually darkened, rest of antennomeres black.

Labrum transverse, laterally covered with several pale setae, anterior margin slightly sinuate. Anterior part of head lustrous, sparsely covered with pale setae. Frontal tubercles very large, subtriangular, distinctly elevated, lustrous. Vertex dull, densely covered with small confused punctures and short pale hairs. Antennae filiform, 0.75 times as long as the body, length ratio of antennomeres: 15-8-14-17-14-14-12-12-11-15.

Pronotum transverse, 2.00-2.15 times broader than it is long, widest at the first third, slightly narrowed anteriad and posteriad, dull, densely covered with small punctures and pale hairs. Surface with two small deep depressions laterally. Anterior margin moderately concave, posterior margin nearly straight, lateral margins slightly rounded. Anterior and posterior margins thinly bordered, lateral margins indistinctly bordered. Anterior angles nearly rectangular, posterior angles obtusely angulate, all angles with distinct tooth bearing long pale seta.

Scutellum short, subtriangular with small dense punctures and short pale hairs, dull.

Elytra parallel, dull. Humeral calli well developed. Elytral surface very densely covered with small confused punctures and short pale hairs. Epipleura distinct, gradually narrowed to apex.

Macropterous.

Ventral surface nearly lustrous, finely punctured and covered with pale hairs. Last ventrite with shallow semicircular incision. Claws bifid.

Basimetatarsomere 1.40 times as long as two following metatarsomeres combined.

Female: Frontal tubercles small. Median spot on pronotum very small, placed near the middle of posterior pronotal margin. Last ventrite entire. Claws appendiculate.

Aedeagus as in Fig. 12.

Distribution

Tanzania.
**Diagnosis**

Owing to very large frontal tubercles in males, *A. similis* resembles *A. nodicornis* (Laboissière, 1922), *A. incisitarsis* (Laboissière, 1922), *A. leontovitchi* (Laboissière, 1940) and *Apophylia demeyeri* Bezdék, 2005. All these species differ from *A. similis* in enlarged fifth antennomere (antennae filiform in *A. similis*), *A. incisitarsis* also in deeply incised first mesotarsomere in male (entire in in *A. similis*).

**Comments**

*A. similis* was described based on 4 females. I had the possibility to study 2 syntypes deposited now in ZMHB. One of them is designated here as lectotype. In unidentified material from MRAC and ZMHB I have found short series including several males. Because the male was not known, redescription of this species is presented.

**Apophylia tarsalis** Laboissière, 1938


**Type Material Examined**


**Distribution**

Ethiopia.

**Comments**

This species was described based on one female from south Ethiopia. The exact determination is very problematic due to only one known female. However, it seems to be related to *Apophylia chloroptera* Thomson, 1858. The characters which can be used for identification could be mainly the small yellow transverse frontal tubercles and almost completely black prosternum (pronotum yellow with three black spots).

Although the original locality label bears the locality name “Marale”, the correct name is “Moyale” as it was published in the original description.

**Apophylia trapezicollis** Laboissière, 1940

**Type Material Examined**


**Distribution**

Eritrea.

**Comments**

*A. trapezicollis* was described based on one female. The holotype is very similar to the females of *A. disconotata*. Females of both species differ in the length ratio of antennomeres 2 and 3. Antennomere 3 is 2.2 times longer than antennomere 2 in the female of *A. disconotata*, but only 1.8 times in the female of *A. trapezicollis*. The central spot on pronotum is larger in the female of *A. disconotata*.

*Apophylia vicina* LABOISSIÈRE, 1940


**Type Material Examined**


**Distribution**

Cameroon.

**Comments**

Similarly as in *A. marginicollis*, the male is unknown, thus the exact identification is problematic. The female holotype is very similar to *A. marginicollis*, but differs in the yellow frontal tubercles.

*Chapuisia scutellaris* (ALLARD, 1889), comb. nov.

*Apophylia scutellaris* ALLARD, 1889: C. R. Soc. Ent. Belg., 33: 71 (key), 74 (sep. 6, 9) (Natal); WEISE 1924: 183; WILCOX 1971: 147.
TYPE MATERIAL EXAMINED


DISTRIBUTION
RSA.

COMMENTS
The number of available specimens was not specified by Allard (1889) in the original description. Both specimens found in ISNB and MNHN must be treated as syntypes, although the syntype from MNHN bears the label “Holotype” added by subsequent curator. Although this species was described as *Apophylia*, it is a typical representative of *Chapuisia*.

*Chapuisia weisei* nom. nov.

*Chapuisia scutellaris* Weise, 1926, Arkiv Zool. 18A(34): 19 (nec *Chapuisia scutellaris* (Allard, 1889)).

COMMENTS
*Apophylia scutellaris* Allard, 1889 was transferred to the genus *Chapuisia* (see above). Thus *Chapuisia scutellaris* Weise, 1926 is a secondary homonym and I propose new name *Chapuisia weisei* for it.

DESCRIPTIONS OF NEW TAXA

*Apophylia grobbelaarae* n. sp.

TYPE MATERIAL


**DESCRIPTION**

Body length of males 3.95-4.60 mm (holotype 4.45 mm); of females 4.50-5.60 mm.

Male. Body flattened, parallel, densely pubescent, semiopaque. Head bicolourous; vertex, upper half of postgenae, frontal tubercles, mandibles and last palpomere black, anterior and ventral parts (including lower half of postgenae) yellow. Antennomeres 1 to 3 yellow, antennomeres 4 and 6 gradually darkened, rest of antennomeres black. Pronotum variable: 1) usually black with somehow paler extreme margins, 2) paler males have yellow pronotum with three large connected black spots, 3) palest males with three well separated black spots. Scutellum and underside black. Elytra metallic green. Legs yellow, femora with basal two thirds black (paler males only with small spot on femoral base), outer sides of tibiae distinctly darkened, last two tarsal segments infuscate. Labrum transverse, laterally covered with several pale setae, anterior margin sinuate. Anterior part of head semiopaque, sparsely covered with pale setae.
Frontal tubercles small, subtriangular, nearly lustrous. Vertex dull, densely covered with confused punctures and short pale setae. Antennae 0.75 times as long as the body, length ratio of antennomeres 1 to 11: 13-7-13-18-14-13-12-12-11-9-13.

Pronotum transverse, 2.00 – 2.05 times broader than it is long, widest at the first third, slightly narrowed anteriad and posteriad, semiopaque, densely covered with small punctures and pale setae. Surface with two feeble depressions laterally. Anterior margin slightly concave, posterior margin straight, lateral margins slightly rounded. Anterior and posterior margins thinly bordered, lateral margins indistinctly bordered. Anterior angles nearly rectangular, posterior angles obtusely angulate, all angles bear long pale seta.

Scutellum short, semicircular with small dense punctures and short pale hairs, dull. Elytra parallel, dull. Humeral calli well developed. Elytral surface very densely covered with small confused punctures and short pale hairs. Epipleura distinct, gradually narrowed to apex.

Macropterous.

Ventral surface lustrous, finely punctured and covered with pale hairs. Last ventrite with semicircular incision. Basimetaatarsomere 1.3 times as long as two following metatarsomeres combined.

Female: Pronotum bicolorous, yellow with well separated three black spots. Legs paler than in males, usually yellow with slightly darkened femoral base and infuscate tarsi. Last ventrite entire. Claws appendiculate.

The shape of aedeagus as in Fig. 13.

**Distribution**

RSA, Zimbabwe.

**Host plant**

According to label data, several specimens were collected feeding on *Ehretia rigida* (Boraginaceae). One paratype was collected by suction machine from *Acacia nilotica* (Mimosoideae) but the feeding was not observed.

**Diagnosis**

Dark males of *A. grobbelaarae* n. sp. are very similar to the males of *A. aurolimbata*, but males of *A. grobbelaarae* n. sp. have yellow underside of head, while it is black in *A. aurolimbata*. Females and paler males of *A. grobbelaarae* n. sp. with yellow pronotum with three black spots resemble *A. clavareau* and *A. lindae* n. sp. and females of *A. aurolimbata*. *A. clavareau* has yellow frontal tubercles (black in *A. grobbelaarae* n. sp.). *A. lindae* n. sp. has more robust yellow antennae and yellow postgenae and abdomen (antennae filiform, postgenae bicolorous and abdomen black in *A. grobbelaarae* n. sp.), females of *A. aurolimbata* have black underside of head (yellow in *A. grobbelaarae* n. sp.). All species mentioned here distinctly differ in the structure of aedeagi (Figs 1-3, 13, 15).
ETYMOLOGY
Dedicated to Elisabeth Grobbelaar (SANC), a specialist in Chrysomelidae, who kindly send me abundant Apophylia material including main part of the type series of A. grobbelaarae n. sp.

Apophylia lesnei aethiopica n. ssp.

TYPE MATERIAL
Holotype (male) and 2 paratypes (1 male, 1 female), labelled: “Harar Prov.: Errer 1200 m [p] 21/22 [h] ,VIII.1971 [w, p] // Coll. Mus. Tervuren Ethiopie R.O.S. Clarke [w, p]“ (in MRAC). The specimens of the newly described species are provided with one red label: „HOLOTPUS [or PARATYPUS], Apophylia lesnei aethiopica s. ssp., det. J. Bezděk 2005”.

DESCRIPTION
Body length of males 7.30-7.60 mm (holotype 7.30 mm); of female 7.85 mm.

Labrum transverse, covered with several pale setae, anterior margin distinctly sinuate. Anterior part of head lustrous, sparsely covered with pale setae. Frontal tubercles small, subtriangular, covered with microsculpture, semiopaque. Interantennal space with small deep groove. Frons with feeble depression behind the frontal tubercles. Vertex dull, densely covered with small confused punctures and short pale hairs. Antennae 0.70 times as long as the body, length ratio of antennomeres 1 to 11: 23-10-17-29-22-19-18-19-16-17-20.

Pronotum transverse, 1.95 times broader than it is long, widest in the middle, slightly narrowed anteriad and posteriad, dull, densely covered with small punctures and pale hairs. Surface with two feeble depressions laterally. Anterior margin slightly concave, posterior margin nearly straight, with indicated shallow incision in the middle, lateral margins slightly rounded. Anterior and posterior margins thinly bordered, lateral margins indistinctly bordered. Anterior angles nearly rectangular, posterior angles obtusely angulate, all angles with small tooth bearing long pale seta.

Scutellum short, subtriangular, with small dense punctures and short pale hairs, semiopaque.

Elytra parallel, dull. Humeral calli well developed. Elytral surface very densely covered with small confused punctures and short pale hairs. Epipleura distinct, gradually narrowed to apex.

Macropterous.

Ventral surface lustrous, finely punctured and covered with pale hairs. Last ventrite with very deep subtrapezoidal incision. First tarsomeres of all tarsi
enlarged. Basimetatarsomere 1.1 times as long as two following metatarsomeres combined (Fig. 8). Claws bifid.

Female: First tarsomeres not enlarged. Last ventrite with small sharp incision. Claws appendiculate.

The shape of aedeagus as in Fig. 5.

**DISTRIBUTION**

Ethiopia.

**DIAGNOSIS**

*A. lesnei aethiopica* n. ssp. differs from the nominate subspecies in strongly enlarged first tarsomeres in male (basimetatarsomeres as in Figs 7-8). The aedeagi of both subspecies are very similar (Figs 4-5).

**ETYMOLOGY**

Named after Ethiopia, where the type series was collected.

*Apophylia haladai* n. sp.

**TYPE MATERIAL**


**DESCRIPTION**

Body length 5.70-5.85 mm (holotype 5.85 mm).

Male. Body flattened, parallel, densely pubescent, semiopaque. Head bicolorous; frontal tubercles, frons, vertex, upper half of postgenae, apical half of mandibles and last palpomere black, anterior and ventral parts of head yellow. Pronotum yellow with three black spots (median and two lateral). Scutellum black, elytra metallic green. Prosternum yellow with middle and anterior parts black. Mesosternum black, mesoepimera yellow. Metasternum black with yellow posterior margin, metaepisterna yellow. Abdomen black. Legs yellow, outer side of tibiae darkened, last two tarsal segments infuscate. Antennomeres 1 to 3 yellow, antennomere 1 with black spot dorsally, antennomeres 4 and 5 yellow ventrally and gradually darkened dorsally, last six antennomeres black.

Labrum transverse, covered with several pale setae, anterior margin sinuate. Anterior part of head nearly lustrous, sparsely covered with pale setae. Frontal tubercles large, subtriangular, lustrous. Vertex dull, densely covered with small confused punctures and short pale hairs. Antennae slender, 0.75 times as long as the body, length ratio of antennomeres 1 to 11: 21-9-15-23-17-16-15-14-12-10-
16. Antennomeres 3 to 5 slightly flattened, ventrally densely covered with long pale setae.

Pronotum transverse, 2.15 times broader than it is long, widest at the first third, slightly narrowed anteriad and posteriad. Anterior margin semiopaque, rest of surface dull, densely covered with small punctures and pale hairs. Surface with two deep depressions laterally. Anterior margin moderately concave, posterior margin nearly straight, lateral margins slightly rounded. Anterior and posterior margins thinly bordered, lateral margins indistinctly bordered. Anterior angles nearly rectangular, posterior angles obtusely angulate, all angles with distinct tooth bearing long pale seta.

Scutellum short, subtriangular with small dense punctures and short pale hairs, dull.

Elytra parallel, dull. Humeral calli well developed. Elytral surface very densely covered with small confused punctures and short pale hairs. Epipleura distinct, gradually narrowed to apex.

Macropterous.

Ventral surface lustrous, finely punctured and covered with pale hairs. Last ventrite with semicircular incision. Tarsi slender. Basimetatarsomere 1.6 times as long as two following metatarsomeres combined.

Female. Last ventrite entire. Claws appendiculate.

The shape of aedeagus as in Fig. 14.

**Distribution**

Guinea.

**Diagnosis**

*Apophila haladai* n. sp. resembles *A. chloroptera* THOMSON, 1858. Males of *A. chloroptera* have deeply incised first mesotarsomere, which is simple in *A. haladai* n. sp.

**Etymology**

Dedicated to Marek HALADA, a specialist in Chrysidae (Hymenoptera), who collected the holotype.

*Apophyla lindae* n. sp.

**Type material**

VAAL MUSEUM [blue label, p]“ (in TMSA); 3 paratypes (males), labelled: “Abachaus [placed in Namibia, Otjiwarongo District], III 53, G. Hobohm [w, p] / collection TRANSVAAL MUSEUM [blue label, p]“ (in TMSA). The specimens of the newly described species are provided with one red label: „HOLOTYPUS [or PARATYPUS], Apophylia lindae n. sp., det. J. Bezděk 2005”.

DESCRIPTION
Body length of males 4.70-6.25 mm (holotype 5.30 mm); of females 6.05-6.75 mm.

Male. Body flattened, parallel, densely pubescent, semiopaque. Head yellow (including postgenae and frontal tubercles), vertex black, mandibles at apices darkened. Pronotum yellow with three black spots (median and two lateral). Scutellum black, elytra metallic green. Prosternum yellow with darkened anterior margin. Mesosternum black, mesoepimera yellow. Metasternum black with yellow posterior margin, metaeepisterna yellow. Abdomen yellow, sometimes with darkened base. Legs yellow, all femora with small black spot on the base (in paler specimens, the spot distinct only on profemora), last two tarsal segments infuscate. Antennae yellow, antennomeres 7 to 11 with black apices (dark specimens with antennomere 7 to 11 completely black).

Labrum transverse, laterally covered with several pale setae, anterior margin distinctly sinuate. Anterior part of head lustrous, sparsely covered with pale setae. Frontal tubercles small, subtriangular, nearly lustrous. Vertex dull, densely covered with small confused punctures and short pale hairs. Antennae slender, but relatively short and robust, 0.65 times as long as the body, length ratio of antennomeres 1 to 11: 16-8-12-19-17-16-13-11-10-14. Antennomeres 3 to 8 slightly flattened.

Pronotum transverse, 1.85-1.90 times broader than it is long, widest at the first third, narrowed anteriad and posteriad, dull, densely covered with small punctures and pale hairs. Surface with two depressions laterally shallowly connected in the middle. Anterior margin moderately concave, posterior margin nearly straight, shallowly concave in the middle. Anterior and posterior margins thinly bordered, lateral margins indistinctly bordered. Anterior angles nearly rectangular, posterior angles nearly rounded, all angles with small tooth bearing long pale seta.


Abdomen semiopaque, densely covered with short pale setae. Meso- and metasternum lustrous, finely punctured and covered with longer pale setae. Last ventrite with deep semicircular incision. Basimetatarsomere 1.2 times as long as two following metatarsomeres combined. Claws bifid.

Female: Last ventrite entire. Claws appendiculate.
The shape of aedeagus as in Fig. 15.
DISTRIBUTION
Namibia.

DIAGNOSIS
A. lindae n. sp. resembles A. clavareau, paler specimens of A. grobbelaarae n. sp. and females of A. aurolimbata, but differs in the combination of the following characters: head yellow including postgenae and frontal tubercles, short and robust antennae almost completely yellow, legs yellow with small black spots on bases of femora. All species mentioned here distinctly differ also in the structure of aedeagi (Figs 1-3, 13, 15).

ETYMOLOGY
Dedicated to Linda VÁVROVÁ, my goddaughter.

Apophylia marketae n. sp.

TYPE MATERIAL
Holotype (male) and 15 paratypes (14 males, 1 female), labelled: “TANZANIA CE, W of Mbuyuni, (E of Iringa), 9.3.2002, M. Snížek leg. [w, p]” (HT and 5 PT in NMPC, rest in JBCB); 30 paratypes (23 males, 7 female), labelled: “Tanzania CE, SE of Mbuyuni, Baobab vall., (NE of Iringa), 9.3.2002, M. Snížek leg. [w, p]” (3 PT in FKCC, 3 PT in NHMB, 3 PT in JVCJ, rest in JBCB); 1 paratype (male), labelled: “ZIMBABWE 11.xii.1993 17°53′S/25°49′E, lux, Victoria Falls: Zambezi-NP-Camp, leg. J. Deckert [blue label, p]” (in ZMHB); 2 paratypes (males), labelled: “ZAMBIA 23.iii.1993 13°06′03″S 31°47′32″E South Luangwa NP, Mfuwe Crocodile Farm, 450m, lux, leg. M. Uhlig [blue label, p]” (in ZMHB). The specimens of the newly described species are provided with one red label: „HOLOTYPUS [or PARATYPUS], Apophylia marketae n. sp., det. J. Bezděk 2005”.

DESCRIPTION
Body length of males 4.75-5.45 mm (holotype 4.80 mm); of females 5.75-6.50 mm.

Male. Body flattened, parallel, densely pubescent, semiopaque. Head yellow, mandibles dark brown to black, vertex with median black spot not touching inner margin of eyes. Pronotum yellow with three black spots (median and two lateral). Scutellum black, elytra metallic green. Underside yellow. Legs yellow, last two tarsal segments infuscate. Antennae yellow with slightly darkened apices of antennomeres. Last two or three antennomeres sometimes completely dark brown.

Labrum transverse, covered with several pale setae, anterior margin distinctly sinuate. Anterior part of head nearly semiopaque, sparsely covered with pale setae. Interantennal space with small groove. Frontal tubercles small, subtriangular, semiopaque, covered with microsculpture. Vertex dull, densely covered with small confused punctures and short pale hairs. Antennae filiform, 0.85 times as long
as the body, length ratio of antennomeres 1 to 11: 14-7-14-19-18-16-15-14-12-10-14.

Pronotum transverse, 1.85-1.95 times broader than it is long, widest at the first third, slightly narrowed anteriad and posteriad. Surface semiopaque, densely

14-17. Aedeagus (a - dorsal view, b - lateral view): 14 - *Apophylia haladai* n. sp., 15 - *A. lindae* n. sp., 16 - *A. marketae* n. sp., 17 - *A. dellacasai* n. sp. Scale 1 mm
covered with small punctures and pale hairs. Surface with two deep depressions laterally. Anterior margin slightly concave, posterior margin nearly straight, lateral margins slightly rounded. Anterior and posterior margins thinly bordered, lateral margins indistinctly bordered. Anterior angles nearly rectangular, posterior angles obtusely angulate, all angles with distinct tooth bearing long pale seta.

Scutellum short, subtriangular, with small dense punctures and short pale hairs, dull.

Elytra parallel, dull. Humeral calli well developed. Elytral surface very densely covered with small confused punctures and relatively long pale hairs. Epipleura distinct, gradually narrowed to apex.

Macropterous.

Ventral surface lustrous, finely punctured and covered with pale hairs. Last ventrite with semicircular incision. Basimetatarsomere 1.5 times as long as two following metatarsomeres combined. Claws bifid.

Female. Last ventrite entire. Claws appendiculate.
The shape of aedeagus as in Fig. 16.

**DISTRIBUTION**

Tanzania, Zambia, Zimbabwe.

**DIAGNOSIS**

Owing to its coloration, filiform antennae and slender tarsi in the male, *A. marketae* can be compared only with *A. marshalli* and *A. jeanneli*. All three species can be exactly identified only based on the structure of aedeagus (Figs 10-11, 16).

**ETYMOLOGY**

Dedicated to Markéta Vávrová, my dear lifelong friend.

**Apophylia dellacasai** n. sp.

**TYPE MATERIAL**

DESCRIPTION

Body length of males 4.45-5.00 mm (holotype 4.95 mm); of females 5.10-6.25 mm.

Male. Body flattened, parallel, densely pubescent, semiopaque. Head yellow, vertex with median black spot usually not touching inner margin of eyes (rarely
vertex completely black or black with small yellow spots behind eyes), apices of mandibles black. Pronotum yellow with three black spots (median and two lateral). Scutellum black. Elytra metallic green, blue or coppery. Underside yellow, median part of metasternum and abdomen rarely darkened. Legs yellow, last two tarsal segments usually infuscate. Antennae yellow, from antennomere 6 gradually darkened (pale specimens with completely yellow antennae).

Labrum transverse, covered with several pale setae, anterior margin very slightly sinuate. Anterior part of head semiopaque, sparsely covered with pale setae. Frontal tubercles small, subtriangular, semiopaque, covered with microsculpture. Interantennal space with shallow groove. Vertex with distinct ridge in the middle, dull, densely covered with small confused punctures and short pale hairs. Antennae slender, 0.65–0.75 times as long as the body, length ratio of antennomeres 1 to 11: 17-8-12-17-14-15-13-12-11-10-12. Antennomeres are relatively short and robust.

Pronotum transverse, 2.05 times broader than it is long, widest at the first third, slightly narrowed anteriad and posteriad. Surface with two deep depressions laterally, densely covered with small punctures and pale hairs, nearly lustrous, depressions semiopaque. Anterior margin moderately concave, posterior margin nearly straight, lateral margins slightly rounded. Anterior and posterior margins thinly bordered, lateral margins indistinctly bordered. Anterior angles nearly rectangular, posterior angles obtusely angulate, all angles with distinct tooth bearing long pale seta.

Scutellum short, subtriangular with small dense punctures and short pale hairs, dull.

Elytra parallel, dull. Humeral calli well developed. Elytral surface very densely covered with small confused punctures and short pale hairs. Epipleura distinct, gradually narrowed to apex.

Macropterous.

Ventral surface lustrous, finely punctured and covered with pale hairs. Last ventrite with semicircular incision. Tarsi short and relatively robust. Basimetatarsomere 1.05 times as long as two following metatarsomeres combined.

Female. Last ventrite entire. Claws appendiculate.

The shape of aedeagus as in Fig. 17.

**Distribution**

Egypt, Oman.

**Diagnosis**

*Apophylia dellacasai* n. sp. somehow resembles *A. marketae* n. sp., *A. marshalli* and *A. jeanneli*, but differs in the short and robust antennae and tarsi and in the structure of aedeagus (Figs 10-11, 16-17).
ETYMOLOGY
Dedicated to Marco DELLACASA (CIUC), a specialist in Aphodiidae, who collected the main part of the type series.

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